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## Poly(2-oxazoline) molecular brushes by grafting through of poly(2-oxazoline)methacrylates with aqueous ATRP

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Molecular brushes of poly(2-oxazoline)s (POx) are an intriguing class of polymers as they combine a unique architecture with the properties of POx as a biomaterial. Here, the synthesis of several POx macromonomers with methacrylate end groups and consecutive grafting through polymerization by aqueous atom transfer radical polymerization (ATRP) at room temperature is reported. <sup>1</sup>H-NMR spectroscopy and size exclusion chromatography (SEC) confirmed the synthesis of POx molecular brushes with maximum side chain grafting densities, narrow molar mass distributions ( $D \leq 1.16$ ) and final molar masses corresponding to the initial macromonomer : initiator ratio. Chain extension experiments show high end group fidelity and formation of block copolymer molecular brushes, and kinetic studies revealed a polymerization behavior of oligo(2-methyl-2-oxazoline) methacrylate very similar to the frequently used oligo(ethylene glycol) methacrylate (OEGMA<sub>475</sub>). Aqueous solutions of POx molecular brushes with poly(2-ethyl- and 2-isopropyl-2-oxazoline) side chains exhibit the typically defined thermoresponsive behavior with a tunable, very narrow and reversible phase transition.

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### Introduction

Molecular brushes are composed of a macromolecular backbone with a maximum number of pending polymeric or oligomeric side chains. The very high grafting density induces side chain as well as backbone stretching, which results in a characteristic elongated, worm-like shape of the molecular brush.<sup>1</sup> The synthesis of molecular brushes is classified into three methods of which every single one has its specific advantages. These methods are *grafting onto* (synthesis of side chains and the backbone with a subsequent coupling), *grafting from* (polymerization of side chains starting from a macroinitiator backbone) and *grafting through* (polymerization of macromonomers).<sup>1</sup> The unique structure, considerable size and high aspect ratio of molecular brushes make them suitable for use as single molecule nanomaterials to be used as templates, actuators and sensors in nano(bio)technology<sup>2–4</sup> or even in nanomedicine as drug-delivery systems.<sup>5,6</sup>

Molecular brushes based on poly(2-oxazoline)s (POx) came into the focus of recent research because of the advantageous properties of POx as a biomaterial. POxs are pseudo-peptides synthesized by living cationic ring-opening polymerization

(LCROP) of 2-oxazolines in a controlled manner ( $D < 1.2$ , adjustable molar masses).<sup>7,8</sup> Similar to poly(ethylene glycol) (PEG), hydrophilic POx is non-toxic<sup>9,10</sup> and non-immunogenic (low to none complement activation)<sup>10,11</sup> and suppresses biofouling,<sup>12,13</sup> POxylated entities display the same “stealth effect” as PEGylated ones,<sup>14–16</sup> and hydrophilic as well as amphiphilic POxs show a biodistribution and excretion which is beneficial for medical applications.<sup>17</sup> Additionally their solubility and aggregation behavior can be fine-tuned by structural and compositional variation of the poly(2-oxazoline)s by the use of easily accessible monomers.<sup>8,18–20</sup> Furthermore, multiple functionalization and structural versatility are possible using respective initiators, monomers and terminating agents.<sup>7,21–30</sup> After early approaches<sup>31,32</sup> yielding a variety of POx-based comb polymers,<sup>33–46</sup> POx molecular brushes have been synthesized recently by several routes. The majority of the applied methods focuses on the polymerization of macromonomers by group transfer polymerization (GTP), free radical polymerization (FRP) or reversible addition/fragmentation chain transfer polymerization (RAFT). GTP and RAFT can result in molecular brushes with narrow molar mass distributions, but are still limited to relatively short backbones and side chains.<sup>31,47,48</sup> Unfortunately, GTP is demanding with respect to the strict reaction conditions and the more robust RAFT has to be stopped at rather low conversions (usually ~50%) in order to suppress side reactions.<sup>48</sup> Nevertheless, RAFT has successfully been used for the synthesis of copolymer molecular brushes

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made from short ( $P_n = 5$ ) 2-ethyl- and 2-nonyl-2-oxazoline macromonomers.<sup>49</sup> FRP of macromonomers results in molecular brushes with a very long backbone ( $P_w \sim 1050$ ) but a broad molar mass distribution.<sup>50</sup> Alternatively, in different grafting from approaches, 2-isopropenyl-2-oxazoline was polymerized using FRP, anionic polymerization or rare earth metal catalyzed GTP to create a backbone from which different 2-oxazolines were polymerized *via* living cationic ring-opening polymerization (LCROP) to yield molecular brushes.<sup>51,52</sup> The anionic polymerization resulted in defined brushes, even with block copolymer side chains, but again under very demanding reaction conditions. The rare earth metal catalyzed GTP is stated to perform better, but only indicated by AFM-imaging so far.<sup>52</sup> In addition, the rare earth metal catalyst is rather difficult to handle. One of the most interesting features of POx molecular brushes is their defined thermoresponsive behavior. The temperature dependent solubility in water can be fine-tuned by varying the length of the backbone, the side chain or the side chain composition.<sup>48,50,51,53–56</sup> Thus, POx molecular brushes are interesting materials to be used as sensors or, with cloud points adjustable close to the human body temperature, for biomedical applications such as theragnostics. Furthermore, it is possible to conjugate antibodies to molecular brushes of 2-ethyl- and 2-isopropyl-2-oxazoline, which could be interesting for the development of active targeting in drug-delivery.<sup>57</sup>

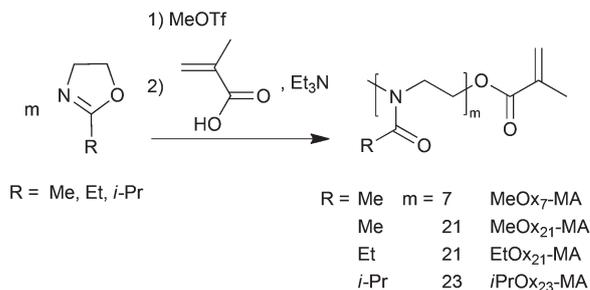
As outlined above, the existing methods for the synthesis of POx molecular brushes are either experimentally demanding or uncontrolled and/or limited to rather short side chain or backbone lengths. Here, the atom transfer radical polymerization (ATRP) should be an attractive alternative to overcome these drawbacks. However, first attempts to use ATRP for the synthesis of POx molecular brushes were made by ATRP of 2-isopropenyl-2-oxazoline and subsequent LCROP of 2-oxazolines, thus a typical grafting from approach. Unfortunately, ATRP of 2-isopropenyl-2-oxazoline yielded only oligomeric products, which has contributed to a strong complexation of the copper species used for the ATRP catalyst by the monomer and the growing oligomers.<sup>55</sup> To the best of our knowledge, ATRP of POx macromonomers has not been attempted so far and could be more viable. For example, a similar oligo(ethylene glycol) methacrylate can be polymerized by aqueous ATRP in a fast and well-controlled manner. Even grafting from a protein is possible.<sup>58,59</sup> Moreover, the synthesis of suitable POx macromonomers is well known and has been reported in the literature.<sup>31,37,48,60–62</sup>

Here we report on the well-controlled synthesis of POx molecular brushes by the grafting through of poly(2-oxazoline) methacrylates by aqueous ATRP.

## Results and discussion

### Synthesis of poly(2-oxazoline)methacrylate macromonomers

Defined POx macromonomers were synthesized according to Kobayashi *et al.*<sup>60</sup> by LCROP of 2-methyl- (MeOx), 2-ethyl-



**Scheme 1** Synthesis of poly(2-oxazoline) macromonomers by LCROP with methyl triflate as the initiator and methacrylic acid/triethylamine termination.

(EtOx) or 2-isopropyl-2-oxazoline (iPrOx) with methyl triflate as the initiator at 90 °C in acetonitrile (Scheme 1). The LCROP was terminated by a mixture of methacrylic acid and triethylamine creating the termination agent, a methacrylate anion, *in situ* to yield the methacrylate end functionalized POx (MeOx<sub>m</sub>-MA, EtOx<sub>m</sub>-MA and iPrOx<sub>m</sub>-MA). Analogous to oligo(ethylene glycol) macromonomers, short POx macromonomers with  $m = 7$  but also longer ones with  $m = 21, 23$  were synthesized to investigate their consecutive polymerizability by ATRP. The length of the macromonomers was controlled by the initial  $[M]_0 : [I]_0$  ratio. While MeOx yields highly water-soluble polymers, EtOx and especially iPrOx give thermosensitive polymers.<sup>63–68</sup>

The four POx-MA macromonomers were analyzed by means of SEC, matrix assisted laser desorption ionization–time-of-flight mass spectrometry (MALDI-ToF-MS) and <sup>1</sup>H-NMR spectroscopy. With the latter, the chemical composition of all macromonomers could be confirmed, especially the methacrylic end function by the appearance of two singlets around 5.8 ppm, corresponding to the two vinylic protons (Fig. 1). Comparison of the peak integral ratio of the terminal methyl group originating from the initiator (two multiplets between 2.8 and 3.1 ppm) with the methacrylic group indicates a quantitative functionalization by the termination reaction. The <sup>1</sup>H-NMR spectrum also shows minor traces of diethyl ether (from precipitation) and triethylammonium ions (from the termination reaction). The impurities could be removed by dialysis, however, because of the relatively low molar mass of the macromonomers; this also caused a significant loss of the product, and since both impurities are not interfering with the consecutive ATRP reaction, the macromonomers were used as such for further experiments. The presence of triethylammonium salt caused obstruction in the SEC trace of the low molar mass MeOx<sub>7</sub>-MA as both compounds had a similar elution time. This resulted in a relatively high apparent dispersity of  $D_{SEC} = 1.31$  while all other macromonomers gave dispersity values of  $D_{SEC} = 1.08–1.09$  and, thus, close to the theoretical limit (Fig. 2a, Table 1). This is also corroborated by the MALDI-ToF-MS data, as only molar mass distributions corresponding to the desired macromonomer structure of poly(2-methyl-2-oxazoline) ( $\Delta m/z = 85$ ), poly(2-ethyl-2-oxazoline)



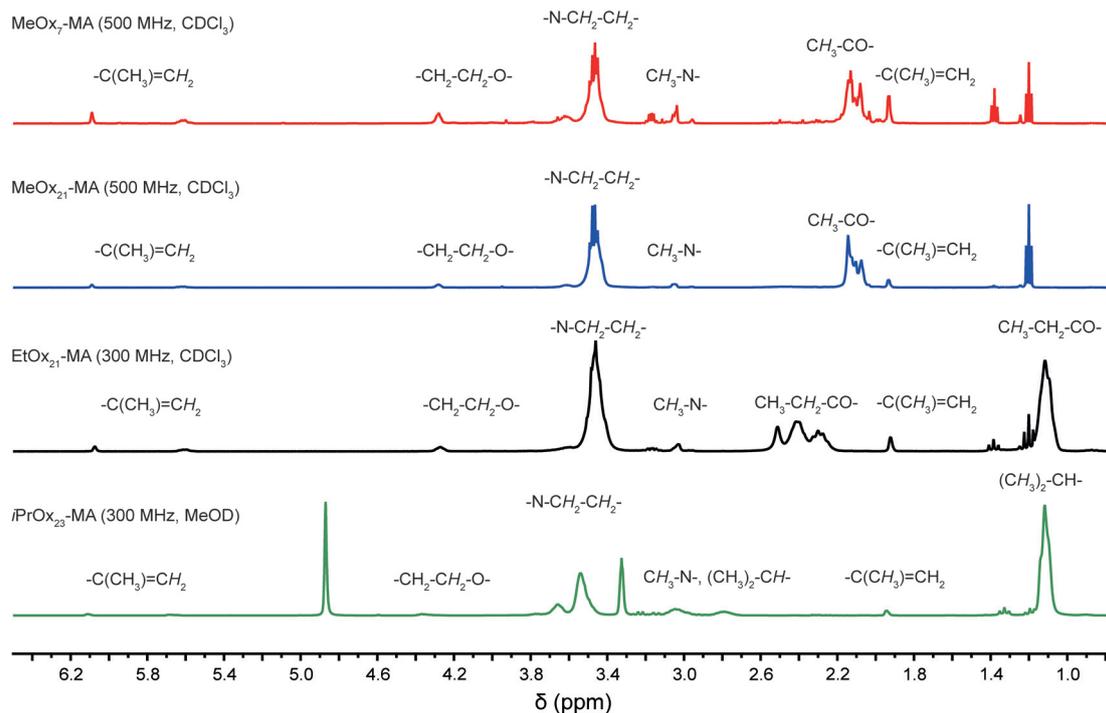


Fig. 1  $^1\text{H-NMR}$ -spectra of the synthesized macromonomers with signals aligned to the respective structural units.

( $\Delta m/z = 99$ ) or poly(2-isopropyl-2-oxazoline) ( $\Delta m/z = 113$ ) with a methyl group at the one and a methacrylic group at the other chain end can be observed in the MALDI-ToF-MS spectra (Fig. 2b and c). A straightforward analysis of the short MeOx<sub>7</sub>-MA is again obstructed only this time, by the overlap of the mass signals of the macromonomer with matrix components. Nevertheless, close inspection of the mass spectrum indicates a much better dispersity of MeOx<sub>7</sub>-MA as calculated from the deformed SEC elugram.

### Synthesis of POx molecular brushes

We tested several ATRP systems with different ligands (*N,N,N',N',N''*-pentamethyldiethylenetriamine (PMDETA), tris[2-(dimethylamino)ethyl]amine (Me<sub>6</sub>TREN), 2,2'-bipyridine (bpy)), with or without initial addition of a deactivating copper(II)-complex to polymerize the POx macromonomers to molecular brushes (Table 2).

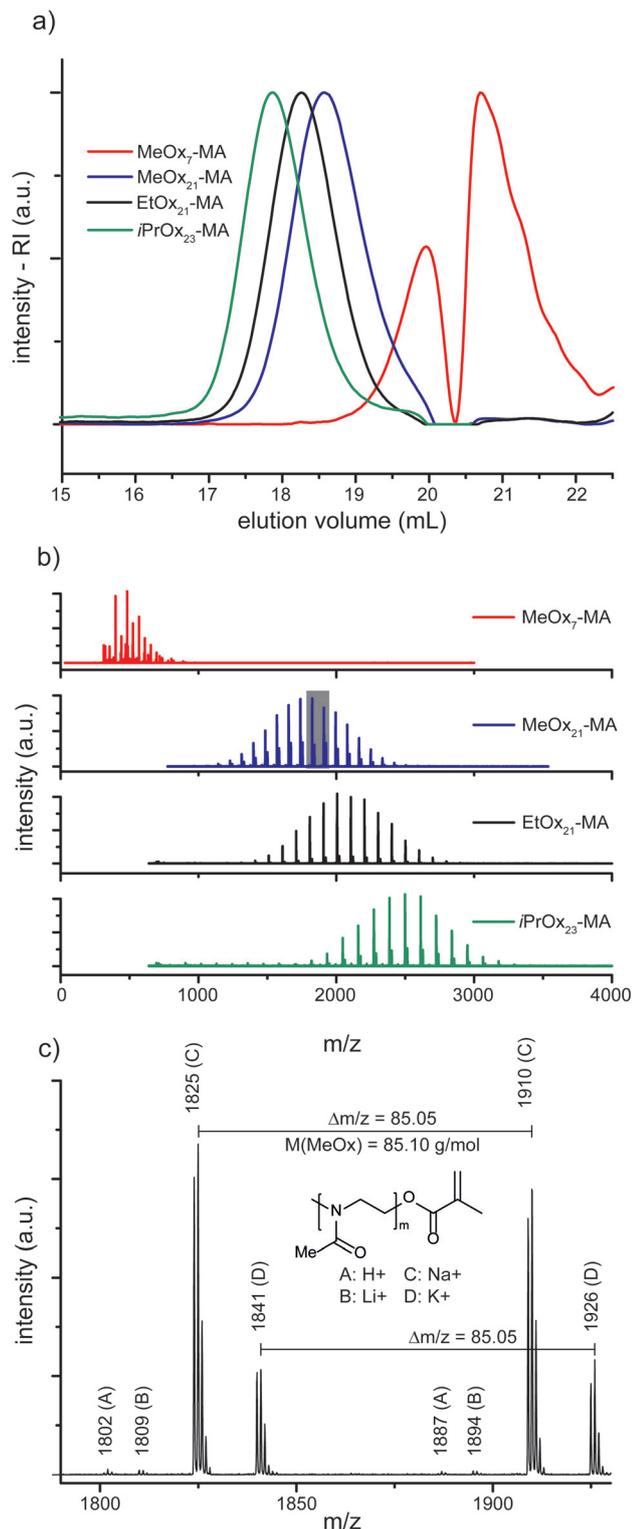
As is obvious from Table 2, entries 1–3, no controlled ATRP reaction was possible using PMDETA, Me<sub>6</sub>TREN or bpy without additional Cu(II) as indicated by the high values for  $D_{\text{SEC}}$  and apparent molar masses being significantly higher than expected. This is in agreement with earlier literature reports. The highly reactive complexes with PMDETA and especially Me<sub>6</sub>TREN disproportionate in aqueous solution into a more complex reaction system that result in high radical concentration and loss of polymerization control. Even when using water stable and less reactive bpy complexes, initially added Cu(II) species might be necessary to maintain reaction control.<sup>58,69–71</sup> Accordingly, the use of 2,2'-bipyridine as the

ligand and copper(I)- as well as copper(II)-bromide (Table 2, entry 4) in aqueous solution proved to be a suitable method and result in the desired molecular brushes in high yields, with narrow molar mass distributions and final molar masses in the expected range (Scheme 2). While the initiator, 2-hydroxyethyl-2-bromoisobutyrate (HEBIB), and copper(I)-complex were used in equimolar amounts, a nine-fold excess of the deactivating copper(II)-complex was necessary to achieve and maintain satisfying reaction control even at high conversions of the macromonomers. A similar recipe was also successfully applied for the polymerization of oligo(ethylene glycol) methacrylate (OEGMA<sub>475</sub>) by ATRP.<sup>58</sup>

A series of eleven molecular brushes was successfully synthesized by using this approach and analyzed by SEC, NMR and SEC-MALLS (Table 3). In general, conversions >80% could be reached within 6 h or less. The residual macromonomer and catalyst could be removed by column chromatography. Fig. 3a shows the SEC elugrams of a series of molecular brushes P(MeOx<sub>7</sub>-MA)<sub>n</sub> with  $n = 10$ –820 and the steady increase of the molar masses and low dispersities ( $D = 1.16$ –1.21) for all brushes. The symmetric SEC traces exhibit neither high nor low molecular subpopulation, which indicates a fast and quantitative initiation, and chain transfer or termination reactions seem to be not significant.

$^1\text{H-NMR}$  spectroscopy verified the anticipated structure of the synthesized brushes. All observable signals can be assigned to the respective structural units as shown exemplarily for P(MeOx<sub>7</sub>-MA)<sub>52</sub> in Fig. 3b. Unfortunately, end group analysis is not possible because of the very low relative inten-





**Fig. 2** Characterization of the POx-MA macromonomers. (a) SEC eluograms of the macromonomers showing a narrow molar mass distribution and the growth of the molar mass with increasing side chain length and degree of polymerization. (b) MALDI-ToF-MS spectra of the macromonomers demonstrating a narrow molar mass distribution and only the desired macromonomer species. Distribution of MeOx<sub>7</sub>-MA is overlapped by peaks of the matrix. (c) Detailed section of the spectrum of MeOx<sub>21</sub>-MA. The chosen part is highlighted (grey area) in (b).

sity of the terminal hydroxyl group and the broad strong proton signals of the high molar mass brush. The two singlets of the vinylic protons of the macromonomers around 5.8 ppm are no longer observable and new broad signals between 0.4 and 1.4 ppm arose, indicating the formation of the methacrylate backbone. Comparing the intensities of these backbone signals with signals originating from the side chain terminal methyl groups around 3.0 ppm (3H), a maximum grafting density can be concluded for the series of P(MeOx<sub>*m*</sub>-MA)<sub>*n*</sub>. This analysis was not possible for P(EtOx<sub>*m*</sub>-MA)<sub>*n*</sub> and P(iPrOx<sub>*m*</sub>-MA)<sub>*n*</sub> because of signal overlapping (data not shown).

Also longer macromonomers (MeOx<sub>21</sub>-MA and EtOx<sub>21</sub>-MA) could be successfully polymerized to molecular brushes and for all cases the dispersity as determined by SEC ( $D_{\text{SEC}}$ ) is very low (Table 3). As can be expected, the number average molar masses of the molecular brushes as determined by SEC deviate from the expected values especially for longer side chain brushes because of the different macromolecular architecture of the used calibration standard being linear PMMA. Nevertheless, an obvious trend is observable with respect to the increasing molar masses of the brushes with the same side chains but increasing backbone length and for comparable backbones but different side chain lengths. While absolute values differ, these trends at least indicate a controlled polymerization and the possibility of adjustable molar masses by ATRP. For a better analysis, we performed additional characterization by SEC with light scattering detection (SEC-MALLS, see below).

Naturally, the realizable backbone length of molecular brushes by the grafting through of longer macromonomers is limited by the viscosity of the initial reaction solution. We found that a simple dilution resulted in a noticeable loss of control of the ATRP system as the higher dispersity and deviation of calculated vs. determined molar masses for P(MeOx<sub>7</sub>-MA)<sub>820</sub> and P(EtOx<sub>21</sub>-MA)<sub>183</sub> shows (Table 3). The latter could only be obtained with a double amount of the activator copper complex. Here, alternative controlled/living radical polymerization techniques such as Cu<sup>0</sup>-mediated living radical polymerizations might be more useful.<sup>72–75</sup> Related studies are currently ongoing. Nevertheless, aqueous ATRP was found to be a very versatile polymerization to obtain highly defined molecular brushes with considerable side chain and backbone lengths. As the aqueous ATRP can be driven to high conversions it represents a good alternative to other CRP techniques such as RAFT.<sup>48</sup> Only the polymerization of iPrOx<sub>23</sub>-MA turned out to be difficult and stopped at a conversion of around 66%. Already the initial reaction solution was very viscous because of the higher molar mass of iPrOx<sub>23</sub>-MA and further significant viscosity increase upon conversion of the macromonomer made stirring impossible.

Further analysis of the POx molecular brushes by SEC-MALLS revealed molar masses that corresponded well with the theoretical values in the majority of the cases and corroborates the assumption of a controlled grafting through polymerization. The traces obtained from SEC-MALLS have a symmetrical appearance and again narrow molar mass distributions



**Table 1** Synthesis details and analytical data of the POx<sub>m</sub>-MA macromonomers

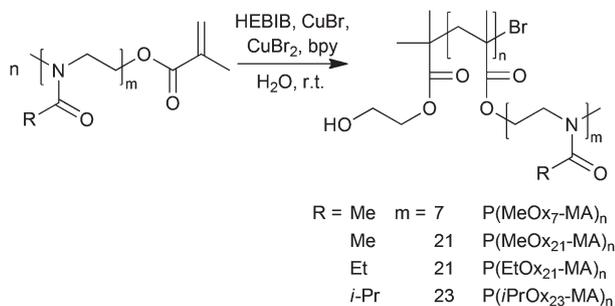
Name	R	[M] <sub>0</sub> /[I] <sub>0</sub>	M <sub>theo.</sub> <sup>a</sup> (g mol <sup>-1</sup> )	P <sub>n,NMR</sub> <sup>b</sup>	M <sub>n,NMR</sub> <sup>b</sup> (g mol <sup>-1</sup> )	M <sub>n,MALDI</sub> <sup>c</sup> (g mol <sup>-1</sup> )	M <sub>n,SEC</sub> <sup>d</sup> (g mol <sup>-1</sup> )	D <sub>SEC</sub> <sup>d</sup>
MeOx <sub>7</sub> -MA	Me	5	525	7	695	524	424	1.31 <sup>e</sup>
MeOx <sub>21</sub> -MA	Me	20	1802	21	1887	1790	2010	1.09
EtOx <sub>21</sub> -MA	Et	20	2083	21	2182	2050	2484	1.08
iPrOx <sub>23</sub> -MA	i-Pr	20	2363	23	2703	2450	3063	1.08

<sup>a</sup> As calculated from [M]<sub>0</sub>: [I]<sub>0</sub>. <sup>b</sup> As determined by end group analysis from <sup>1</sup>H-NMR spectroscopy data. <sup>c</sup> As determined by MALDI-ToF-MS. <sup>d</sup> As determined by SEC (DMAc, 5 g L<sup>-1</sup> LiBr, 1 vol% H<sub>2</sub>O, calibrated with PMMA standards). <sup>e</sup> Apparent dispersity obtained from the obstructed elution peak as shown in Fig. 2a. The actual dispersity is significantly lower.

**Table 2** Experimental conditions for the ATRP of MeOx<sub>7</sub>-MA in aqueous solution at room temperature with different ligands and Cu(II) ratios. The reaction time was set to 2 h, and the aimed degree of polymerization was 30

No.	Ligand	[I] (mM)	Conv. <sup>a</sup> (%)	[M]: [I]: [Cu(I)]: [Cu(II)]: [L]	M <sub>n,theo</sub> <sup>b</sup> (kg mol <sup>-1</sup> )	M <sub>n,SEC</sub> <sup>c</sup> (kg mol <sup>-1</sup> )	D <sub>SEC</sub> <sup>c</sup>
1	PMDETA	38	86.7	28 : 1 : 1 : 0 : 2	16.8	23.6	2.83
2	Me <sub>6</sub> TREN	35	89.5	30 : 1 : 1 : 0 : 1	18.8	57.8	2.23
3	bpy	35	93.5	30 : 1 : 1 : 0 : 2	20.0	164	1.41
4	bpy	8	>99	30 : 1 : 1 : 9 : 22	20.7	10.2	1.15

<sup>a</sup> As determined by SEC analysis of the unpurified molecular brush. <sup>b</sup> As calculated from initial macromonomer–initiator ratio and conversion. <sup>c</sup> As determined by SEC, DMAc, 5 g L<sup>-1</sup> LiBr, 1 vol% H<sub>2</sub>O, calibrated with PMMA standards.

**Scheme 2** Synthesis of POx molecular brushes by aqueous ATRP at room temperature from POx macromonomers. The components were typically used at a ratio of [I] : [Cu(I)] : [Cu(II)] : [bpy] = 1 : 1 : 9 : 22.

can be found. Dispersities range even below ( $D \leq 1.07$ ) the values determined by SEC with refractive index detection, probably because of the lower scattering of the lower molar mass fraction. The molar mass of P(iPrOx<sub>23</sub>-MA)<sub>37</sub> could not be determined because of interaction of the polymer with the stationary phase at 35 °C (standard conditions) as well as 25 °C. Higher temperatures could not be used because of the cloud point of the polymer. Unfortunately, also the longest brush P(MeOx<sub>7</sub>-MA)<sub>820</sub> could not be analyzed by SEC-MALLS due to the low amount of polymer product after the work-up procedure, which had to be performed to quantitatively remove the remaining macromonomer and the catalyst. A typical SEC-MALLS trace along with RI detection of P(MeOx<sub>7</sub>-MA)<sub>52</sub> is shown in Fig. 4. The determined molar masses are in

good agreement with the theoretical values. For most cases, the deviation of the determined number average molar masses from the theoretical values is only about 10% and thus within the experimental error of the method (temperature dependence of  $dn/dc$  and underestimation of species with low scattering intensity). The determined molar masses for the longest molecular brushes (P(MeOx<sub>7</sub>-MA)<sub>202</sub> and P(EtOx<sub>21</sub>-MA)<sub>183</sub>) deviate by about 30%, which cannot be accounted for by an uncontrolled ATRP conversion as the dispersity of the products is still very low. One reason could be the strong hygroscopic nature of the used macromonomers, which made it difficult to adjust the correct [M]<sub>0</sub>: [I]<sub>0</sub> ratio. However, during attempts to additionally freeze-dry the educts prior to use, considerable autopolymerization of the macromonomers was observed which forced us to refrain from additional drying steps. The SEC-MALLS elugram in Fig. 4a shows an additional distribution at very high molar masses which is not appearing in the refractive index detection. Similar elugrams with high molar mass distributions but of strongly varying intensities were also observed for other P(MeOx<sub>7</sub>-MA)<sub>n</sub> molecular brushes and are attributed to very large metastable brush aggregates. First dynamic light scattering measurements in methanol (data not shown) revealed only one particle distribution assigned to the molecular brushes, underlining the assumption of aggregates only stabilized by hydrophobic interactions in aqueous solution.

Investigation of the polymerization kinetics of MeOx<sub>7</sub>-MA as the POx analogous to the frequently used OEGMA<sub>475</sub> revealed a very similar polymerization behavior of both macromonomers (Fig. 5) under comparable conditions. For MeOx<sub>7</sub>-MA as well as for OEGMA<sub>475</sub> a similar deviation from ideal



**Table 3** Experimental and analytical data of the molecular brushes synthesized by aqueous ATRP with [I] : [Cu(I)] : [Cu(II)] : [bpy] = 1 : 1 : 9 : 22; reaction times are as listed

Name	$n^a$	[I] (mM)	$t_R$ (h)	Conv. <sup>b</sup> (%)	$M_{n,theo}^a$ (kg mol <sup>-1</sup> )	$M_{n,SEC}^c$ (kg mol <sup>-1</sup> )	$D_{SEC}^c$	$M_{n,SEC-MALLS}^d$ (kg mol <sup>-1</sup> )
P(MeOx <sub>7</sub> -MA) <sub>10</sub>	10	5	2	>99	7.4	6.9	1.16	— <sup>f</sup>
P(MeOx <sub>7</sub> -MA) <sub>52</sub>	52	5		>99	36.1	15.5	1.16	40.0
P(MeOx <sub>7</sub> -MA) <sub>104</sub>	104	5		>99	72.3	24.2	1.16	65.9
P(MeOx <sub>7</sub> -MA) <sub>202</sub>	202	5		>99	140.5	36.3	1.15	101.1
P(MeOx <sub>7</sub> -MA) <sub>820</sub>	820	1	3	82.1	695.0	93.5	1.21	— <sup>f</sup>
P(MeOx <sub>21</sub> -MA) <sub>50</sub>	50	5	2	86.3	95.1	42.3	1.08	95.7
P(MeOx <sub>21</sub> -MA) <sub>91</sub>	91	2.5	6	86.6	172.4	55.9	1.09	150.1
P(EtOx <sub>21</sub> -MA) <sub>50</sub>	50	5	2	89.2	108.2	48.6	1.06	118.4
P(EtOx <sub>21</sub> -MA) <sub>92</sub>	92	2.5	6	86.0	200.6	71.9	1.08	212.6
P(EtOx <sub>21</sub> -MA) <sub>183</sub>	183	1 <sup>e</sup>	5	90.9	398.7	84.4	1.10	276.6
P(iPrOx <sub>23</sub> -MA) <sub>37</sub>	37	5	2	65.4	99.2	46.4	1.09	— <sup>f</sup>

<sup>a</sup> As calculated from the initial macromonomer–initiator ratio and conversion. <sup>b</sup> As determined by SEC analysis of the unpurified molecular brush. <sup>c</sup> As determined by SEC, DMAc, 5 g L<sup>-1</sup> LiBr, 1 vol% H<sub>2</sub>O, calibrated with PMMA standards. <sup>d</sup> As determined by SEC-MALLS, H<sub>2</sub>O, 0.72 g L<sup>-1</sup> NaN<sub>3</sub>, 8.5 g L<sup>-1</sup> NaNO<sub>3</sub>, 35 °C. <sup>e</sup> [I] : [Cu(I)] : [Cu(II)] : [bpy] = 1 : 2 : 9 : 25. <sup>f</sup> Not determined due to the low amount of polymer or because of polymer–column interactions.

pseudo-first order kinetics is observable.<sup>58</sup> ATRP of MeOx<sub>7</sub>-MA resulted in a linear increase of the molar mass as a function of the polymerization time at low dispersities ( $D \leq 1.18$ ) even at high conversions.

Chain extension of a molecular brush was successfully performed and further corroborates the assumption of a controlled polymerization of POx macromonomers without a significant loss of chain end functionality. First, MeOx<sub>21</sub>-MA was polymerized to a conversion of >90%, an aliquot of the reaction solution was collected and analyzed and the remaining solution was further converted *in situ* by the addition of EtOx<sub>21</sub>-MA to give a P[(MeOx<sub>21</sub>-MA)-(EtOx<sub>21</sub>-MA)] block copolymer brush. SEC analysis of the products reveals very low dispersities of both polymers, especially for the block copolymer with a complete shift to a higher molar mass with no indication of the residual homopolymer brush (Fig. 6). Even though the remaining MeOx<sub>21</sub>-MA participates in the second polymerization, this indicates high end group fidelity and demonstrates the possibility of preparing block copolymer molecular brushes by the grafting through of POx macromonomers by aqueous ATRP.

### Thermoresponsive behavior

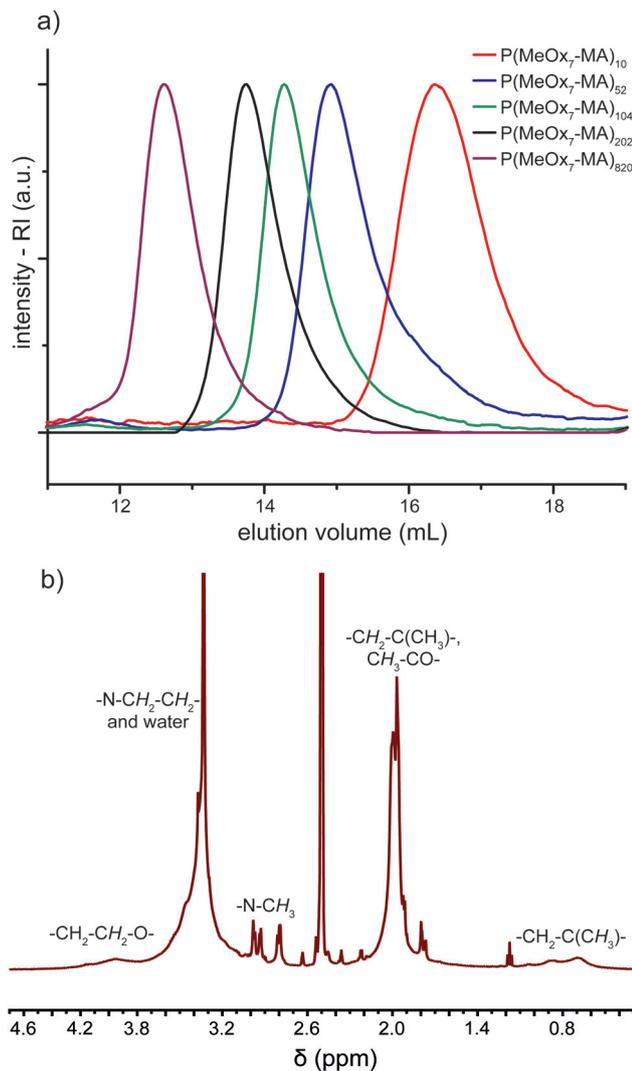
The synthesized molecular brushes based on poly(2-ethyl-2-oxazoline) and poly(2-isopropyl-2-oxazoline) should exhibit a thermoresponsive behavior similar to earlier observations.<sup>48,50,51,53–56</sup> The transmittance of aqueous solutions (1 wt%) of the brushes was observed in dependence of the temperature and the cloud point ( $T_{CP}$ ) was determined at 10% decrease in optical transmittance (Fig. 7). All four brushes showed very sharp transitions that were in the range of only 1–1.5 K. During several consecutive cooling/heating cycles, the materials showed no deviation of the  $T_{CP}$  and only a minor heating–cooling hysteresis of approximately 1 K as

shown for P(EtOx<sub>21</sub>-MA)<sub>50</sub> and P(iPrOx<sub>23</sub>-MA)<sub>37</sub> in Fig. 7. The determined cloud points appeared in the expected range of  $T_{CP} = 73$  °C for P(EtOx<sub>21</sub>-MA)<sub>50</sub> and  $T_{CP} = 36$  °C for P(iPrOx<sub>23</sub>-MA)<sub>37</sub>. An increase of the molar mass of P(EtOx<sub>21</sub>-MA)<sub>n</sub> resulted in the expected slight decrease of  $T_{CP}$ .<sup>48,51,55</sup> The phase transition of P(iPrOx<sub>23</sub>-MA)<sub>37</sub> is intriguingly close to the human body temperature and falls nicely into the row of previously determined  $T_{CP}$ 's of P(iPrOx<sub>m</sub>)<sub>n</sub> molecular brushes, as a brush made by consecutive living anionic polymerization (backbone) and LCROP (grafting from of the side chains) with  $n = 52, 132, 216$  and  $m = 24$  showed  $T_{CP} = 31, 29.1$  and  $27$  °C, respectively.<sup>55</sup>

## Conclusion

A series of molecular brushes of poly(2-oxazoline)s was synthesized by grafting through polymerization of oligo- and poly(2-oxazoline) macromonomers by aqueous ATRP at room temperature. These molecular brushes feature high grafting densities, narrow molar mass distributions and adjustable molar masses. It was found that surprisingly long and high molar mass molecular brushes are accessible by the grafting through approach; however, the method is limited by the initial viscosity of the reaction solution. Oligo(2-methyl-2-oxazoline) methacrylate showed a very similar polymerization behavior to the commercially available OEGMA<sub>475</sub> including the deviation from first order kinetics as well as low dispersities and almost linear increase of the molar mass with the ATRP reaction time. Thus, a versatile POx-based macromonomer to OEGMA<sub>475</sub> is now available for the synthesis of structurally complex biomaterials. Chain extension and formation of block copolymer molecular brushes were also successful. Aqueous solutions of POx molecular brushes with poly(2-ethyl- and 2-isopropyl-2-oxazoline) side chains exhibit the typically defined thermore-





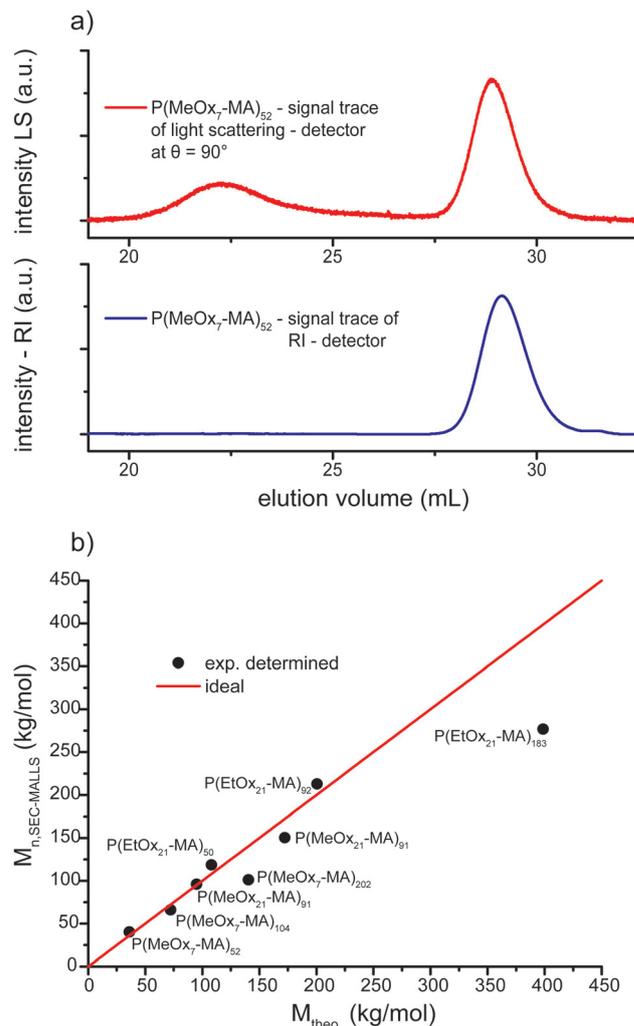
**Fig. 3** (a) SEC elugrams (DMAc, 5 g L<sup>-1</sup> LiBr, 1 vol% H<sub>2</sub>O, calibrated with PMMA standards) of five molecular brushes from MeOx<sub>7</sub>-MA with increasing backbone length. (b) <sup>1</sup>H-NMR-spectrum (500 MHz, DMSO-d<sub>6</sub>) of P(MeOx<sub>7</sub>-MA)<sub>52</sub> with assignment of the respective structural units.

sponsive behavior with a tunable, very narrow and reversible phase transition. The  $T_{CP}$  of P(iPrOx<sub>23</sub>-MA)<sub>37</sub> lies at 36 °C close to the human body temperature and falls nicely into the list of previously reported P(iPrOx<sub>m</sub>)<sub>n</sub> molecular brushes synthesized by grafting from and living ionic polymerization. In conclusion, aqueous ATRP of oligo- and poly(2-oxazoline) methacrylates was found to be a versatile synthetic route for the preparation of defined POx molecular brushes.

## Experimental

### Materials and methods

All chemicals were purchased from Sigma-Aldrich (Steinheim, Germany) or Acros Organics (Geel, Belgium) and were used as received unless stated otherwise. 2-Methyl-, 2-ethyl-, 2-iso-

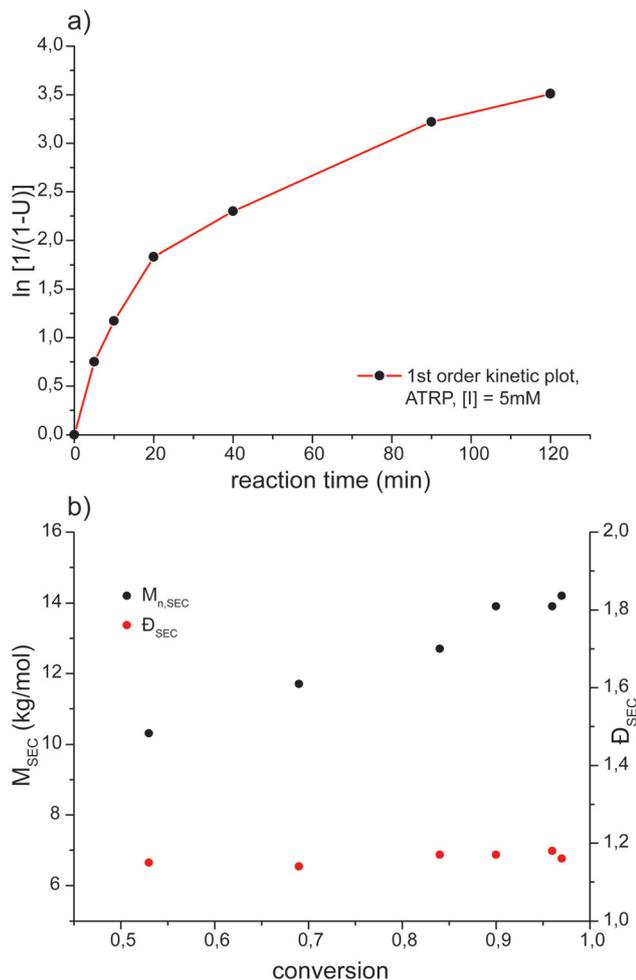


**Fig. 4** (a) SEC-MALLS elugram of P(MeOx<sub>7</sub>-MA)<sub>52</sub> (H<sub>2</sub>O, 0.72 g L<sup>-1</sup> NaN<sub>3</sub> and 8.5 g L<sup>-1</sup> NaNO<sub>3</sub>, 35 °C) in red along with SEC-RI detection (blue). The distribution of the molecular brushes is visible at elution volumes between 27.5 and 31 mL with a narrow and symmetrical appearance. The second high molar mass peak is attributed to metastable brush aggregates. (b) Comparison of the determined number average molar masses with the theoretical values as calculated from the initial macro-monomer/initiator ratio and the conversion. Ideal behavior is indicated by the red line. Only P(MeOx<sub>7</sub>-MA)<sub>202</sub> and P(EtOx<sub>21</sub>-MA)<sub>183</sub> deviate noticeably.

propyl-2-oxazoline, methyltriflate and triethylamine were dried by refluxing over calcium hydride and by distillation under a dry nitrogen atmosphere. 2-Isopropyl-2-oxazoline was synthesized as described elsewhere.<sup>64</sup> Methacrylic acid was distilled *in vacuo* to remove the inhibitor and stored under a dry nitrogen atmosphere at 7 °C. Aqueous solutions were prepared with deionized water (Millipore, 18.2 MΩ cm) unless stated otherwise.

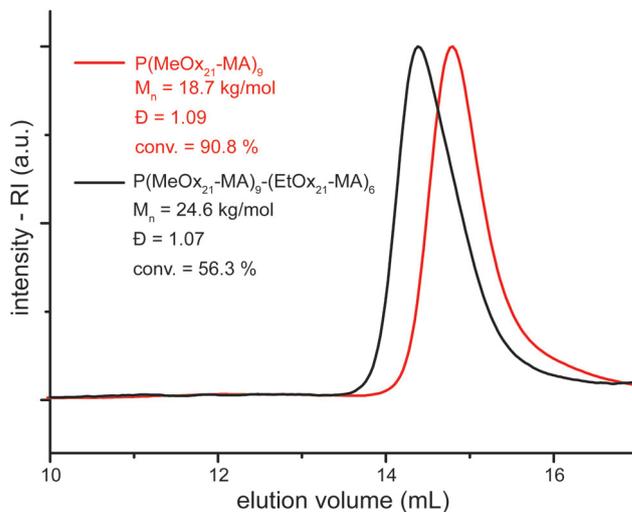
<sup>1</sup>H-NMR-spectroscopy was performed on a DRX 500 P or an AC 300 P (Bruker, Germany), and spectra were calibrated to the solvent residual signal. MALDI-ToF-MS was performed on a Bruker Biflex (Bruker Daltonics, Bremen, Germany) with an



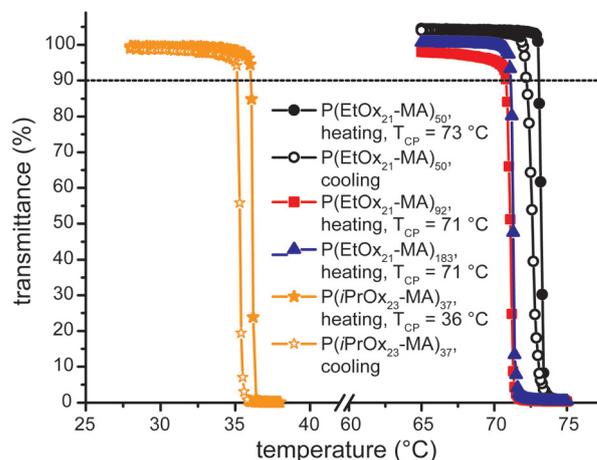


**Fig. 5** (a) First order kinetic plot for the synthesis of molecular brushes based on poly(2-methyl-2-oxazoline) at room temperature in aqueous solution. Initiator concentration  $[I] = 5 \text{ mM}$ ;  $[\text{macromonomer}] : [I] : [\text{Cu}^{\text{II}}] : [\text{bpy}] = 50 : 1 : 1 : 9 : 22$ . (b) Development of  $M_{n,\text{SEC}}$  and  $D_{\text{SEC}}$  with the conversion.

$\text{N}_2$ -laser ( $\lambda = 337 \text{ nm}$ ) in positive reflector mode. Spectra were recorded in a range of 40–4400 Da and the matrix was suppressed up to 330 Da (MeOx<sub>7</sub>-MA), 450 Da (EtOx<sub>21</sub>-MA, IPOx<sub>23</sub>-MA) or 500 Da (MeOx<sub>21</sub>-MA), respectively. The device was calibrated from 750 to 3150 Da with Peptide Calibration Standard II (Bruker) prior to every measurement. Samples were prepared by the dried droplet method. Macromonomers were dissolved at  $1 \text{ g L}^{-1}$  in methanol with 1% trifluoroacetic acid and subsequently mixed in a ratio of 5 : 1 (v/v) with the matrix ( $20 \text{ g L}^{-1}$  sinapinic acid in methanol with 1% trifluoroacetic acid). Turbidity measurements were performed on a Lambda 35 UV/VIS spectrometer equipped with a PTP-1 Peltier System (all from PerkinElmer, Germany) and controlled using Templab software provided by the instrument supplier. The  $T_{\text{CP}}$  of the molecular brushes were determined by spectrophotometric detection of the changes in transmittance at  $\lambda = 500 \text{ nm}$  of 1.0 wt% aqueous solutions. The heating/cooling rate was  $1.0 \text{ K min}^{-1}$ . Given values for the  $T_{\text{CP}}$  were determined as the temp-



**Fig. 6** SEC elugrams of the two reaction stages of the chain extension. First, the brush was synthesized by aqueous ATRP of MeOx<sub>21</sub>-MA with initiator concentration  $[I] = 10 \text{ mM}$ ;  $[\text{MM}]/[I]/[\text{Cu}^{\text{II}}]/[\text{bpy}] = 10/1/1/9/22$ . After 1.5 h, a part of the reaction mixture was transferred into a solution of EtOx<sub>21</sub>-MA for *in situ* chain extension. SEC traces demonstrate a clear shift of the molar mass and the dispersity remains low.



**Fig. 7** Turbidity measurements for the thermoresponsive brushes in aqueous solution (1 wt%). The heating/cooling rate was  $1 \text{ K min}^{-1}$ . For optical clarity, cooling traces are only shown for P(EtOx<sub>21</sub>-MA)<sub>50</sub> and P(iPrOx<sub>23</sub>-MA)<sub>37</sub>, but heating–cooling hysteresis was small ( $\sim 1 \text{ K}$ ) for all brushes.  $T_{\text{CP}}$  was determined at 10% decrease in transmittance on heating.

erature corresponding to a 10% decrease in optical transmittance. Size exclusion chromatography (SEC) was done on a PL-GPC 120, equipped with two Gram 100  $10 \mu\text{L} \times 300 \text{ mm}$  columns and dimethylacetamide (DMAc) with  $5 \text{ g L}^{-1}$  LiBr and 1 vol%  $\text{H}_2\text{O}$  as the mobile phase at  $70 \text{ }^\circ\text{C}$ . The system was calibrated with PMMA standards (PSS, Mainz, Germany) and RI detection. Samples were dissolved in the mobile phase and filtered through  $0.2 \mu\text{m}$  PTFE syringe filters prior to the measurement. SEC-MALLS was performed on a system from Jasco



**Table 4** Results of the  $dn/dc$  measurements with the molecular brushes

Name	$dn/dc$ (mL g <sup>-1</sup> )
P(MeOx <sub>7</sub> -MA) <sub>52</sub>	0.118
P(MeOx <sub>7</sub> -MA) <sub>104</sub>	0.116
P(MeOx <sub>7</sub> -MA) <sub>202</sub>	0.114
P(MeOx <sub>21</sub> -MA) <sub>50</sub>	0.146
P(MeOx <sub>21</sub> -MA) <sub>91</sub>	0.153
P(EtOx <sub>21</sub> -MA) <sub>50</sub>	0.149
P(EtOx <sub>21</sub> -MA) <sub>92</sub>	0.156
P(EtOx <sub>21</sub> -MA) <sub>183</sub>	0.153

(Gross-Umstadt, Germany) with a PU 2080 HPLC-pump, a Jet-Stream II Plus column oven equipped with one SUPREMA 10  $\mu$ m 100 Å  $8 \times 300$  mm column and two SUPREMA 10  $\mu$ m 3000 Å  $8 \times 300$  mm columns (all from PSS). As the mobile phase, double distilled water with 0.72 g L<sup>-1</sup> NaN<sub>3</sub> and 8.5 g L<sup>-1</sup> NaNO<sub>3</sub> was used at a temperature of 35 °C. A Dawn DSP laser photometer at  $\lambda = 632.8$  nm (Wyatt Technology, Dernbach, Germany) and an RI-930 RI detector (Jasco) were used for detection. Samples were dissolved in the mobile phase and filtered through 0.22  $\mu$ m CME-filters prior to the measurement. Refractive index increments  $dn/dc$  were determined on a differential refractometer DR1/b from SLS Systemtechnik (Denzlingen, Germany) in the concentration range of 1–5 g L<sup>-1</sup> at a temperature of 25 °C. Polymers were weighed immediately after lyophilization and dissolved in double distilled water containing 0.72 g L<sup>-1</sup> NaN<sub>3</sub> and 8.5 g L<sup>-1</sup> NaNO<sub>3</sub>. Results of the  $dn/dc$  measurement can be found in Table 4.

### Macromonomer synthesis

**Macromonomer MeOx<sub>7</sub>-MA.** In a glove box, 5.7774 g (35.2 mmol, 1 eq.) methyltriflate was dissolved in 45 mL acetonitrile and 14.9237 g (175.4 mmol, 5.0 eq.) 2-methyl-2-oxazoline were added slowly at r.t. The solution was stirred at 90 °C for 20 min. Afterwards the solution was cooled to r.t. and 7.6507 g (88.9 mmol, 2.5 eq.) methacrylic acid and 8.9936 g (88.9 mmol, 2.5 eq.) triethylamine were added. The solution was stirred at 70 °C overnight. After cooling to r.t., approx. 3 g K<sub>2</sub>CO<sub>3</sub> were added and the mixture was stirred for 24 h. The resulting suspension was centrifuged, and the solution was decanted and filtered through a 20  $\mu$ m PTFE syringe filter. The solvent was removed *in vacuo* and the residual solid was cleaned by three-fold precipitation from methanol in a 10-fold excess of ice-cold dry diethyl ether. After several hours of drying under reduced pressure the macromonomer was obtained as a slightly yellow powder (11.7125 g, 63%). <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 1.92 (s, 3 H, CH<sub>3</sub>-C(=CH<sub>2</sub>-)), 2.10 (br, 21 H, CH<sub>3</sub>-CO-), 2.95 and 3.05 (m, 3 H, CH<sub>3</sub>-N-), 3.54 (br, 26 H, -N-CH<sub>2</sub>-CH<sub>2</sub>-), 4.27 (br, 2 H, -CH<sub>2</sub>-CH<sub>2</sub>-O-), 5.60 (s, 1 H, -C=CH<sub>2</sub>), 6.08 (s, 1 H, -C=CH<sub>2</sub>);  $M_{n,NMR} = 695$  g mol<sup>-1</sup>,  $f_{NMR} = 1$ . SEC:  $D_{SEC} = 1.31$ ,  $M_{n,SEC} = 424$  g mol<sup>-1</sup>. MALDI-ToF-MS:  $D_{MALDI} = 1.04$ ,  $M_{n,MALDI} = 524$  g mol<sup>-1</sup>.

**Macromonomer MeOx<sub>21</sub>-MA.** MeOx<sub>21</sub>-MA was obtained as described above using 0.9668 g (5.9 mmol, 1 eq.) methyltriflate in 30 mL acetonitrile, 10.0592 g (118.2 mmol, 20.0 eq.) 2-methyl-2-oxazoline, 1.3253 g (15.4 mmol, 2.6 eq.) methacrylic acid and 1.5622 g (15.4 mmol, 2.6 eq.) triethylamine. The polymerization time was 30 min, and termination occurred overnight. The macromonomer was received as a slightly yellow powder (8.9415 g, 84%). <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 1.92 (s, 3 H, CH<sub>3</sub>-C(=CH<sub>2</sub>-)), 2.09 (br, 63 H, CH<sub>3</sub>-CO-), 2.94 and 3.04 (m, 3 H, CH<sub>3</sub>-N-), 3.51 (br, 82 H, -N-CH<sub>2</sub>-CH<sub>2</sub>-), 4.27 (br, 2 H, -CH<sub>2</sub>-CH<sub>2</sub>-O-), 5.60 (s, 1 H, -C=CH<sub>2</sub>), 6.08 (s, 1 H, -C=CH<sub>2</sub>);  $M_{n,NMR} = 1887$  g mol<sup>-1</sup>,  $f_{NMR} = 1$ . SEC:  $D_{SEC} = 1.09$ ,  $M_{n,SEC} = 2010$  g mol<sup>-1</sup>. MALDI-ToF-MS:  $D_{MALDI} = 1.02$ ,  $M_{n,MALDI} = 1790$  g mol<sup>-1</sup>.

**Macromonomer EtOx<sub>21</sub>-MA.** As described above, EtOx<sub>21</sub>-MA was obtained using 0.7705 g (4.7 mmol, 1 eq.) methyltriflate in 30 mL acetonitrile, 9.3522 g (94.3 mmol, 20.1 eq.) 2-ethyl-2-oxazoline, 1.0241 g (11.9 mmol, 2.5 eq.) methacrylic acid and 1.1956 g (11.8 mmol, 2.5 eq.) triethylamine. The polymerization time was 90 min, and termination was performed overnight. The macromonomer was received as a white powder (7.9394 g, 81%). <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 1.12 (br, 63 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 1.91 (s, 3 H, CH<sub>3</sub>-C(=CH<sub>2</sub>-)), 2.36 (br, 42 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 2.95 and 3.02 (m, 3 H, CH<sub>3</sub>-N-), 3.50 (br, 82 H, -N-CH<sub>2</sub>-CH<sub>2</sub>-), 4.26 (br, 2 H, -CH<sub>2</sub>-CH<sub>2</sub>-O-), 5.58 (s, 1 H, -C=CH<sub>2</sub>), 6.06 (s, 1 H, -C=CH<sub>2</sub>);  $M_{n,NMR} = 2182$  g mol<sup>-1</sup>,  $f_{NMR} = 1$ . SEC:  $D_{SEC} = 1.09$ ,  $M_{n,SEC} = 2484$  g mol<sup>-1</sup>. MALDI-ToF-MS:  $D_{MALDI} = 1.02$ ,  $M_{n,MALDI} = 2050$  g mol<sup>-1</sup>.

**Macromonomer iPrOx<sub>23</sub>-MA.** As described above, iPrOx<sub>23</sub>-MA was obtained using 0.7238 g (4.5 mmol, 1 eq.) methyltriflate in 30 mL acetonitrile, 10.2399 g (90.5 mmol, 20.1 eq.) 2-isopropyl-2-oxazoline, 0.9690 g (11.3 mmol, 2.5 eq.) methacrylic acid and 1.1388 g (11.3 mmol, 2.5 eq.) triethylamine. The polymerization time was 120 min, and termination occurred overnight. The macromonomer was received as a white powder (5.1543 g, 55%). <sup>1</sup>H-NMR (300 MHz, MeOD):  $\delta$  (ppm) = 1.20 (br, 138 H, (CH<sub>3</sub>)<sub>2</sub>-CH-), 2.00 (s, 3 H, CH<sub>3</sub>-C(=CH<sub>2</sub>-)), 2.84 and 3.08 (br, 26 H, CH<sub>3</sub>-N- and (CH<sub>3</sub>)<sub>2</sub>-CH-), 3.70 (br, 90 H, -N-CH<sub>2</sub>-CH<sub>2</sub>-), 4.42 (br, 2 H, -CH<sub>2</sub>-CH<sub>2</sub>-O-), 5.74 (s, 1 H, -C=CH<sub>2</sub>), 6.16 (s, 1 H, -C=CH<sub>2</sub>);  $M_{n,NMR} = 2702$  g mol<sup>-1</sup>,  $f_{NMR} = 1$ . SEC:  $D_{SEC} = 1.08$ ,  $M_{n,SEC} = 3063$  g mol<sup>-1</sup>. MALDI-ToF-MS:  $D_{MALDI} = 1.02$ ,  $M_{n,MALDI} = 2450$  g mol<sup>-1</sup>.

### ATRP of MeOx<sub>7</sub>-MA with different ligands and Cu(II) ratios

In general, MeOx<sub>7</sub>-MA, 2-hydroxyethyl-2-bromoisobutyrate, and the ligand were dissolved in 2 mL of water in a small tube in the desired ratio. The tube was sealed with a rubber septum and the solution was degassed using a nitrogen flow for 30 min. CuBr and CuBr<sub>2</sub> were filled into another tube, sealed and evacuated/purged with dry nitrogen (3 times). Subsequently, the macromonomer, initiator, and ligand solution was transferred to the copper salts using a gas-tight syringe. The reaction mixture was stirred for 2 h at r.t. and after that diluted with water and lyophilized. The resulting colored powders were analyzed by SEC. For further details refer to Table 2.



### Synthesis of molecular brushes (P(Ox<sub>n</sub>-MA)<sub>n</sub>)

**Molecular brushes of MeOx<sub>7</sub>-MA (P(MeOx<sub>7</sub>-MA)<sub>n</sub>).** In a typical run for the synthesis of P(MeOx<sub>7</sub>-MA)<sub>10</sub>, 72.6 mg (0.1045 mmol, 10.4 eq.) MeOx<sub>7</sub>-MA were dissolved in 2 mL of a 5 mM aqueous solution of the initiator 2-hydroxyethyl-2-bromoisobutyrate (2.1105 mg, 0.01 mmol, 1 eq.) in a small tube and sealed with a rubber septum. The solution was degassed using a nitrogen flow for 30 min. Meanwhile 1.5 mg (0.0105 mmol, 1.1 eq.) CuBr, 20.5 mg (0.0918 mmol, 9.2 eq.) CuBr<sub>2</sub> and 34.6 mg (0.2215 mmol, 22.2 eq.) bpy were weighed in another tube and sealed with a rubber septum. Subsequently the tube was carefully evacuated and backfilled with nitrogen three times. Afterwards the macromonomer–initiator solution was transferred to the solid components using a gas-tight syringe. The resulting mixture was stirred at r.t. After 2 h, the reaction was stopped by dilution with oxygen containing Millipore water and the solution was lyophilized. A small sample was taken from the obtained blue powder to determine the conversion by SEC. The residual solid was dissolved in Millipore water again and passed over a short silica column to remove the catalyst and residual macromonomer. The clear, colorless solution was lyophilized once more and the molecular brush was obtained as a slight yellowish powder (41.9 mg, 56%). Conversion was determined from the ratio of the macromonomer- to molecular brush-band-area in the SEC-traces. Conv. >99%. <sup>1</sup>H-NMR (500 MHz, DMSO-d<sub>6</sub>): δ (ppm) = 1.04 (br, 4 H, (CH<sub>3</sub>)<sub>2</sub>C- and -CH<sub>2</sub>-C(CH<sub>3</sub>)-CH<sub>2</sub>-), 1.90 (br, 23 H, -CH<sub>2</sub>-C(CH<sub>3</sub>- and CH<sub>3</sub>-CO-), 2.80 (d) and 2.94 (dd) (3 H, CH<sub>3</sub>-N-), 3.36 (br, 60 H, -N-CH<sub>2</sub>-CH<sub>2</sub>- and water). SEC: *D*<sub>SEC</sub> = 1.16, *M*<sub>n,SEC</sub> = 6.9 kg mol<sup>-1</sup>.

**Molecular brushes of MeOx<sub>21</sub>-MA (P(MeOx<sub>21</sub>-MA)<sub>n</sub>).** Molecular brushes of MeOx<sub>21</sub>-MA were synthesized according to the procedure described above using MeOx<sub>21</sub>-MA as a macromonomer. <sup>1</sup>H-NMR (500 MHz, DMSO-d<sub>6</sub>) exemplary for P(MeOx<sub>21</sub>-MA)<sub>50</sub>: δ (ppm) = 1.04 (br, 3 H, (CH<sub>3</sub>)<sub>2</sub>C- and -CH<sub>2</sub>-C(CH<sub>3</sub>)-CH<sub>2</sub>-), 1.92 (br, 60 H, -CH<sub>2</sub>-C(CH<sub>3</sub>- and CH<sub>3</sub>-CO-), 2.80 (d) and 2.94 (dd) (3 H, CH<sub>3</sub>-N-), 3.35 (br, 84 H, -N-CH<sub>2</sub>-CH<sub>2</sub>- and water).

**Molecular brushes of EtOx<sub>21</sub>-MA (P(EtOx<sub>21</sub>-MA)<sub>n</sub>).** Molecular brushes of EtOx<sub>21</sub>-MA were synthesized according to the procedure described above using EtOx<sub>21</sub>-MA as a macromonomer. <sup>1</sup>H-NMR (500 MHz, DMSO-d<sub>6</sub>) exemplary for P(EtOx<sub>21</sub>-MA)<sub>50</sub>: δ (ppm) = 1.12 (br, 63 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 2.19 (br, 40 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 2.81 (s) and 2.94 (d) (3 H, CH<sub>3</sub>-N-), 3.35 (br, 91 H, -N-CH<sub>2</sub>-CH<sub>2</sub>- and water).

**Molecular brush of iPrOx<sub>23</sub>-MA (P(iPrOx<sub>23</sub>-MA)<sub>n</sub>).** The molecular brush of iPrOx<sub>23</sub>-MA was synthesized according to the procedure described above using iPrOx<sub>23</sub>-MA as a macromonomer. <sup>1</sup>H-NMR (500 MHz, DMSO-d<sub>6</sub>) P(iPrOx<sub>23</sub>-MA)<sub>37</sub>: δ (ppm) = 0.96 (br, 138 H, (CH<sub>3</sub>)<sub>2</sub>-CH-), 2.12 (br, 2 H, -CH<sub>2</sub>-C(CH<sub>3</sub>)-), 2.78 (br, 27 H, (CH<sub>3</sub>)<sub>2</sub>-CH-), 3.00 (d, 3 H, CH<sub>3</sub>-N-), 3.35 (br, 102 H, -N-CH<sub>2</sub>-CH<sub>2</sub>- and water).

Interestingly, the work-up procedure for all molecular brushes was facile as the residual macromonomers as well as the remaining catalyst could be easily and quantitatively removed by column chromatography.

Characterization of molecular brushes was performed by SEC, SEC-MALLS and <sup>1</sup>H-NMR spectroscopy. For a summary of the experimental conditions and analytical data please refer to Table 3.

### Kinetic investigations

The sample for the kinetic investigation was prepared as described above for the synthesis of the molecular brushes. The initiator concentration was 5 mmol L<sup>-1</sup>, MeOx<sub>7</sub>-MA was used as a macromonomer and the reaction was conducted at r.t. Samples of 0.05 mL were taken periodically under an inert atmosphere at *t* = 0, 5, 10, 20, 40, 60, 90 and 120 min and immediately freeze-dried. Conversion, *M*<sub>n,SEC</sub> and *D*<sub>SEC</sub> were determined for every sample using SEC.

### Chain extension experiments

In a glass tube, 198.7 mg (0.1053, 10.5 eq.) MeOx<sub>21</sub>-MA was dissolved in 1 mL of a 10 mM aqueous solution of 2-hydroxyethyl-2-bromoisobutyrate (2.1105 mg, 0.010 mmol, 1 eq.). The tube was sealed with a rubber septum and the solution was purged with nitrogen for 30 min. Meanwhile, 1.4 mg (0.0098 mmol, 1 eq.), CuBr, 20.1 mg (0.0899 mmol, 9.0 eq.), and CuBr<sub>2</sub> and 34.7 mg (0.2222 mmol, 22.2 eq.) bpy were filled into a tube, sealed and evacuated/purged with dry nitrogen (3 times). Subsequently, the macromonomer–initiator solution was added to the solid components with a gas-tight syringe under Schlenk conditions. The reaction mixture was stirred for 1.5 h at r.t. An aliquot of 0.1 mL was taken to determine the conversion, molar mass and dispersity of the first reaction step (conv. = 90.8%, *M*<sub>n,SEC</sub> = 18.7 kg mol<sup>-1</sup>, *D*<sub>SEC</sub> = 1.09). From the remaining solution, 0.8 mL were transferred to a degassed solution of 199.1 mg (0.0912 mmol, 11.4 eq.) EtOx<sub>21</sub>-MA in 0.9 mL water. Immediately afterwards a sample of 0.1 mL was taken. The remaining mixture was stirred for another 3 h. The work-up procedure was as described above. The copolymer brush was obtained as a white powder (183.5 mg, 51%). Conv. = 56.3%. <sup>1</sup>H-NMR (500 MHz, DMSO-d<sub>6</sub>): δ (ppm) = 0.95 (br, 19 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 1.91 (30 H, -CH<sub>2</sub>-C(CH<sub>3</sub>- and CH<sub>3</sub>-CO-), 2.29 (br, 12 H, CH<sub>3</sub>-CH<sub>2</sub>-CO-), 2.81 (m) and 2.94 (d) (3 H, CH<sub>3</sub>-N-), 3.34 (br, 73 H, -N-CH<sub>2</sub>-CH<sub>2</sub>- and water). SEC: *D*<sub>SEC</sub> = 1.07, *M*<sub>n,SEC</sub> = 24.6 kg mol<sup>-1</sup>.

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### References

- 1 S. S. Sheiko, B. S. Sumerlin and K. Matyjaszewski, *Prog. Polym. Sci.*, 2008, **33**, 759–785.
- 2 C. Li, N. Gunari, K. Fischer, A. Janshoff and M. Schmidt, *Angew. Chem., Int. Ed.*, 2004, **43**, 1101–1104.



- 3 C. Tang, B. Dufour, T. Kowalewski and K. Matyjaszewski, *Macromolecules*, 2007, **40**, 6199–6205.
- 4 P. Zhao, L. Liu, X. Feng, C. Wang, X. Shuai and Y. Chen, *Macromol. Rapid Commun.*, 2012, **33**, 1351–1355.
- 5 Y. Geng, P. Dalhaimer, S. Cai, R. Tsai, M. Tewari, T. Minko and D. E. Discher, *Nat. Nanotechnol.*, 2007, **2**, 249–255.
- 6 M. Müllner, S. J. Dodds, T.-H. Nguyen, D. Senyschyn, C. J. H. Porter, B. J. Boyd and F. Caruso, *ACS Nano*, 2015, **9**, 1294–1304.
- 7 K. Aoi and M. Okada, *Prog. Polym. Sci.*, 1996, **21**, 151–208.
- 8 M. Barz, R. Luxenhofer, R. Zentel and M. J. Vicent, *Polym. Chem.*, 2011, **2**, 1900–1918.
- 9 M. Bauer, S. Schroeder, L. Tauhardt, K. Kempe, U. S. Schubert and D. Fischer, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 1816–1821.
- 10 R. Luxenhofer, G. Sahay, A. Schulz, D. Alakhova, T. K. Bronich, R. Jordan and A. V. Kabanov, *J. Controlled Release*, 2011, **153**, 73–82.
- 11 T. X. Viegas, M. D. Bentley, J. M. Harris, Z. Fang, K. Yoon, B. Dizman, R. Weimer, A. Mero, G. Pasut and F. M. Veronese, *Bioconjugate Chem.*, 2011, **22**, 976–986.
- 12 B. L. Farrugia, K. Kempe, U. S. Schubert, R. Hoogenboom and T. R. Dargaville, *Biomacromolecules*, 2013, **14**, 2724–2732.
- 13 N. Zhang, T. Pompe, I. Amin, R. Luxenhofer, C. Werner and R. Jordan, *Macromol. Biosci.*, 2012, **12**, 926–936.
- 14 R. Konradi, B. Pidhatika, A. Mühlebach and M. Textor, *Langmuir*, 2008, **24**, 613–616.
- 15 M. C. Woodle, C. M. Engbers and S. Zalipsky, *Bioconjugate Chem.*, 1994, **5**, 493–496.
- 16 S. Zalipsky, C. B. Hansen, J. M. Oaks and T. M. Allen, *J. Pharm. Sci.*, 1996, **85**, 133–137.
- 17 F. C. Gaertner, R. Luxenhofer, B. Blechert, R. Jordan and M. Essler, *J. Controlled Release*, 2007, **119**, 291–300.
- 18 T. B. Bonné, K. Lüdtkke, R. Jordan and C. M. Papadakis, *Macromol. Chem. Phys.*, 2007, **208**, 1402–1408.
- 19 R. Hoogenboom, F. Wiesbrock, M. A. M. Leenen, H. M. L. Thijs, H. Huang, C.-A. Fustin, P. Guillet, J.-F. Gohy and U. S. Schubert, *Macromolecules*, 2007, **40**, 2837–2843.
- 20 R. Luxenhofer, A. Schulz, C. Roques, S. Li, T. K. Bronich, E. V. Batrakova, R. Jordan and A. V. Kabanov, *Biomaterials*, 2010, **31**, 4972–4979.
- 21 S. Cesana, J. Auernheimer, R. Jordan, H. Kessler and O. Nuyken, *Macromol. Chem. Phys.*, 2006, **207**, 183–192.
- 22 M. W. M. Fijten, C. Haensch, B. M. van Lankvelt, R. Hoogenboom and U. S. Schubert, *Macromol. Chem. Phys.*, 2008, **209**, 1887–1895.
- 23 R. Jordan, K. Martin, H. J. Räder and K. K. Unger, *Macromolecules*, 2001, **34**, 8858–8865.
- 24 S. Kobayashi and H. Uyama, *J. Polym. Sci., Part A: Polym. Chem.*, 2002, **40**, 192–209.
- 25 K. Lava, B. Verbraeken and R. Hoogenboom, *Eur. Polym. J.*, 2015, **65**, 98–111.
- 26 K. Lüdtkke, R. Jordan, P. Hommes, O. Nuyken and C. A. Naumann, *Macromol. Biosci.*, 2005, **5**, 384–393.
- 27 R. Luxenhofer, M. Bezen and R. Jordan, *Macromol. Rapid Commun.*, 2008, **29**, 1509–1513.
- 28 R. Luxenhofer and R. Jordan, *Macromolecules*, 2006, **39**, 3509–3516.
- 29 M. Reif and R. Jordan, *Macromol. Chem. Phys.*, 2011, **212**, 1815–1824.
- 30 C. Taubmann, R. Luxenhofer, S. Cesana and R. Jordan, *Macromol. Biosci.*, 2005, **5**, 603–612.
- 31 Y. Shimano, K. Sato and S. Kobayashi, *Polym. J.*, 1999, **31**, 458–463.
- 32 Y. Shimano, K. Sato and S. Kobayashi, *Polym. J.*, 1999, **31**, 219–225.
- 33 G. David, V. Alupeii, B. C. Simionescu, S. Dincer and E. Piskin, *Eur. Polym. J.*, 2003, **39**, 1209–1213.
- 34 M. Grasmüller, J. C. Rueda-Sanchez, B. I. Voit and O. Nuyken, *Macromol. Symp.*, 1998, **127**, 109–114.
- 35 A. Gross, G. Maier and O. Nuyken, *Macromol. Chem. Phys.*, 1996, **197**, 2811–2826.
- 36 B. Guillermin, V. Darcos, V. Lapinte, S. Monge, J. Coudane and J.-J. Robin, *Chem. Commun.*, 2012, **48**, 2879–2881.
- 37 S. Kobayashi, M. Kaku, S. Sawada and T. Saegusa, *Polym. Bull.*, 1985, **13**, 447–451.
- 38 S. Kobayashi, Y. Shimano and T. Saegusa, *Polym. J.*, 1991, **23**, 1307–1315.
- 39 O. Nuyken, J. Rueda-Sanchez and B. Voit, *Polym. Bull.*, 1997, **38**, 657–664.
- 40 O. Nuyken, J. R. Sanchez and B. Voit, *Macromol. Rapid Commun.*, 1997, **18**, 125–131.
- 41 J. Rueda, S. Zschoche, H. Komber, D. Schmaljohann and B. Voit, *Macromolecules*, 2005, **38**, 7330–7336.
- 42 Y. Shimano, K. Sato, D. Fukui, Y. Onodera and Y. Kimura, *Polym. J.*, 1999, **31**, 296–302.
- 43 S.-I. Shoda, E. Masuda, M. Furukawa and S. Kobayashi, *J. Polym. Sci., Part A: Polym. Chem.*, 1992, **30**, 1489–1494.
- 44 P. Trivedi and D. Schulz, *Polym. Bull.*, 1980, **3**, 37–44.
- 45 H. Uyama, Y. Honda and S. Kobayashi, *J. Polym. Sci., Part A: Polym. Chem.*, 1993, **31**, 123–128.
- 46 R. Weberskirch, R. Hettich, O. Nuyken, D. Schmaljohann and B. Voit, *Macromol. Chem. Phys.*, 1999, **200**, 863–873.
- 47 M. M. Bloksma, C. Weber, I. Y. Perevyazko, A. Kuse, A. Baumgärtel, A. Vollrath, R. Hoogenboom and U. S. Schubert, *Macromolecules*, 2011, **44**, 4057–4064.
- 48 C. Weber, C. R. Becer, R. Hoogenboom and U. S. Schubert, *Macromolecules*, 2009, **42**, 2965–2971.
- 49 C. Weber, M. Wagner, D. Baykal, S. Hoepfner, R. M. Paulus, G. Festag, E. Altuntas, F. H. Schacher and U. S. Schubert, *Macromolecules*, 2013, **46**, 5107–5116.
- 50 J. Bühler, S. Muth, K. Fischer and M. Schmidt, *Macromol. Rapid Commun.*, 2013, **34**, 588–594.
- 51 N. Zhang, S. Huber, A. Schulz, R. Luxenhofer and R. Jordan, *Macromolecules*, 2009, **42**, 2215–2221.
- 52 N. Zhang, S. Salzinger, B. S. Soller and B. Rieger, *J. Am. Chem. Soc.*, 2013, **135**, 8810–8813.
- 53 C. Weber, A. Krieg, R. M. Paulus, H. M. L. Lambermont-Thijs, C. R. Becer, R. Hoogenboom and U. S. Schubert, *Macromol. Symp.*, 2011, **308**, 17–24.



- 54 C. Weber, S. Rogers, A. Vollrath, S. Hoepfener, T. Rudolph, N. Fritz, R. Hoogenboom and U. S. Schubert, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 139–148.
- 55 N. Zhang, R. Luxenhofer and R. Jordan, *Macromol. Chem. Phys.*, 2012, **213**, 973–981.
- 56 N. Zhang, R. Luxenhofer and R. Jordan, *Macromol. Chem. Phys.*, 2012, **213**, 1963–1969.
- 57 J. Bühler, S. Gietzen, A. Reuter, C. Kappel, K. Fischer, S. Decker, D. Schäffel, K. Koynov, M. Bros, I. Tubbe, *et al.*, *Chem. – Eur. J.*, 2014, **20**, 12405–12410.
- 58 S. Averick, A. Simakova, S. Park, D. Konkolewicz, A. J. D. Magenau, R. A. Mehl and K. Matyjaszewski, *ACS Macro Lett.*, 2012, **1**, 6–10.
- 59 X.-S. Wang and S. P. Armes, *Macromolecules*, 2000, **33**, 6640–6647.
- 60 S. Kobayashi, E. Masuda, S. Shoda and Y. Shimano, *Macromolecules*, 1989, **22**, 2878–2884.
- 61 M. Miyamoto, K. Naka, M. Tokumizu and T. Saegusa, *Macromolecules*, 1989, **22**, 1604–1607.
- 62 C. Weber, R. C. Becer, A. Baumgaertel, R. Hoogenboom and U. S. Schubert, *Des. Monomers Polym.*, 2009, **12**, 149–165.
- 63 R. Hoogenboom, H. M. L. Thijs, M. J. H. C. Jochems, B. M. van Lankvelt, M. W. M. Fijten and U. S. Schubert, *Chem. Commun.*, 2008, 5758–5760.
- 64 S. Huber and R. Jordan, *Colloid Polym. Sci.*, 2008, **286**, 395–402.
- 65 P. Lin, C. Clash, E. M. Pearce, T. K. Kwei and M. A. Aponte, *J. Polym. Sci., Part B: Polym. Phys.*, 1988, **26**, 603–619.
- 66 J.-S. Park and K. Kataoka, *Macromolecules*, 2006, **39**, 6622–6630.
- 67 F. Rehfeldt, M. Tanaka, L. Pagnoni and R. Jordan, *Langmuir*, 2002, **18**, 4908–4914.
- 68 C. Weber, R. Hoogenboom and U. S. Schubert, *Prog. Polym. Sci.*, 2012, **37**, 686–714.
- 69 H. Bergenudd, G. Coullerez, M. Jonsson and E. Malmström, *Macromolecules*, 2009, **42**, 3302–3308.
- 70 W. Tang and K. Matyjaszewski, *Macromolecules*, 2006, **39**, 4953–4959.
- 71 N. V. Tsarevsky, T. Pintauer and K. Matyjaszewski, *Macromolecules*, 2004, **37**, 9768–9778.
- 72 B. M. Rosen and V. Percec, *Chem. Rev.*, 2009, **109**, 5069–5119.
- 73 W. Wang, J. Zhao, N. Zhou, J. Zhu, W. Zhang, X. Pan, Z. Zhang and X. Zhu, *Polym. Chem.*, 2014, **5**, 3533–3546.
- 74 T. Zhang, Y. Du, F. Müller, I. Amin and R. Jordan, *Polym. Chem.*, 2015, **6**, 2726–2733.
- 75 Y. Zhang, Y. Wang, C. Peng, M. Zhong, W. Zhu, D. Konkolewicz and K. Matyjaszewski, *Macromolecules*, 2012, **45**, 78–86.

